



ASHI Proficiency Testing Program

2026 HT-2 Survey Instructions

Ship date: September 16, 2026

Result Submission due date: October 23, 2026

(Changes since the 2026 HT-1 survey are highlighted.)

Note: Additional information pertaining to this survey is available in the current [ASHI PT Operations Manual](#).

Survey Description: The ASHI HLA Typing (HT) Survey is designed to assess laboratory performance in HLA typing by any method. The survey consists of two shipments per year, each comprised of five whole blood specimens in CPDA-I anticoagulant. Included in this Survey, the HT samples can also be used to demonstrate proficiency for the detection of HLA-B27 antigen or alleles in whole blood samples. Participants should HLA type the specimens using the method(s) that provide the highest resolution results, or the method most frequently used if multiple methods provide the same level of typing resolution. Laboratories that clinically use more than one method are expected to correlate results twice per year (if PT specimens are used for correlation, this must be done after the survey has closed).

The HT survey allows the reporting of both low-resolution and high-resolution molecular results. Analytes in the HT survey are defined as:

- **Low-resolution HLA typing (HLA Class I and Class II)**
- **High-resolution HLA typing (HLA Class I and Class II)**
by molecular methods.

Please note that beginning in 2026, HLA Class I and HLA Class II typing results are combined into a single analyte and are entered together.

Laboratories must specify the method used to type each sample and record the manufacturer/kit information. Methods include: SSO, SSP, RT-PCR (Real Time PCR), NGS, Nanopore, PacBio and “Other”.

Analytes in B27 survey are defined as **B27-positive** or **B27-negative**. Laboratories are requested to specify the technology utilized.

Addressing CMS Regulations: Laboratories that are subject to Centers for Medicare and Medicaid Services (CMS) regulation must refer to the most recent guidance document released by the Accreditation Review Board (ARB) regarding proficiency testing of samples and analytes when multiple methods are used. Laboratories are encouraged to contact their ARB commissioners or CMS with questions regarding compliance with CMS proficiency testing regulations.

Deadline: For the 2026 HT-2 survey, the deadline for submitting results is October 23, 2026 at 11:59 PM, Pacific Time (38 days after the shipment date). The specific due date is listed in the PT Shipping Schedule posted on the ASHI PT website and in the ASHI PT Data Center. Late submission of results will not be accepted.

Online Entry of Results: Results must be entered online at <https://ashi-pt.learningbuilder.com/>

A pre-registered user ID and password are required. If you have forgotten your password, follow the on-screen instructions to have it sent to your e-mail address.

To enter HT/B27 Survey results:

1. Select the “PT Results” tab.
2. Locate the correct year and shipment (Spring: HT-1; Fall: HT-2).
3. Click the orange “Begin” button.
4. On the “Awaiting Results” screen, click “Begin Survey.”

Low resolution and high resolution HLA typing analytes will be pre-selected. If your laboratory does not wish to be graded on certain analytes, remove those that you do not wish to be graded on by clicking the “x” next to the analyte name.

If you are reporting DPBI results, you must select the high-resolution HLA typing analyte.

Select the method used for the HT survey from the drop-down menu and enter the Manufacturer/Kit information. Select the B27 method from its drop-down menu.

If different methods were used for different specimens, select the most commonly used method. You may edit this information for individual specimens later.

Click “Continue” to proceed.

Specimen Quality: Specimen quality will default to “Good” for all specimens.

To change specimen quality:

1. Click the three dots [...] next to “Saved.”
2. Select “Change Quality.”
3. Click “OK.”
4. Select the appropriate quality from the drop-down menu.
5. Click “Save.”

If “Acceptable” or “Unacceptable” is selected, describe the issue in the Comment field.

All samples marked “Unacceptable” will automatically be assigned NT1 (not tested due to sample quality), and you will not be able to enter results for these samples.

If a specimen was not tested for a reason other than sample quality, select:

- NT2 (not tested due to a reagent problem or technical failure), or
- NT6 (not tested due to other reasons),

and describe the issue in the Comment field.

If any NT code is selected, results cannot be entered for that specimen.

If only certain loci in a specimen could not be tested, please see below in the HT Test Results Entry section for instructions.

Specimen Quality Definitions

- **Good** - Specimens produced reliable results with no processing or testing problems
- **Acceptable** - Specimens presented difficulties but reliable results were obtained.
- **Unacceptable** - Specimens were unsuitable for testing or produced invalid/unreliable results due to specimen condition.

HT TEST RESULT ENTRY

Before You Begin

- You must complete Sample Quality entry for all specimens before entering HT results.
- Results are grouped into:
 - HT Typing (Class I and Class II combined)
 - B27 (only present for participants enrolled in the B27 survey)
- Confirm that you have selected all analytes you want graded.
 - To change analytes: click the three dots (“...”) beside “Awaiting Result Entry” (top right) → select “Change Analytes.”

Entering Results

1. Click “Enter results” for the first specimen.
2. To change the Test Method:
 - Click “x”
 - Select a new method from the drop-down
 - Manually enter the manufacturer and kit name(s)
3. Enter HLA Typing Results for each locus (HLA class I and HLA class II)
 - Results may be entered at:
 - Low resolution (1-field)
 - High resolution (minimum 2-field)
 - Grading resolution is based on your selected analytes.
 - Typing results beyond 2 fields are not graded (but may appear in summary reports for educational purposes).
 - Low-resolution typing must be sufficient to assign serologic splits. Some alleles may require 2-field genotype results to satisfy this requirement. A list of serological splits can be accessed at: <https://hla.alleles.org/>

General Entry Rules

- Do **not** repeat the locus name in the result field.
- Select the locus name for DRB3/4/5.
- Report alleles/antigens in **numerical order**.
- Use only:
 - Numerals
 - Colons
 - Valid suffixes: G, L, N, P, Q, S
- Use correct WHO nomenclature (e.g., A*01:04:01:01N should not be reported as A*01:04N and A*01:01:01G should not be reported as A*01:01G).
- For each locus:
 - Report two alleles at the same resolution, or
 - Leave both fields blank.
- If a locus was not typed:
 - Leave fields blank.
 - Provide the reason in the Verification tab “Comments” field.

Reporting Homozygous Results:

All loci except DRB3/4/5:

- If only one allele is detected, report it **twice**.
- Do **not** use dashes (“-”) when reporting homozygous typing results.

DRB3/4/5 Special Rules:

- If DRB1 supports DRB3/4/5 homozygosity:
 - Report the allele twice.
 - e.g. when the detected alleles are homozygous DRB1*11:01 and homozygous DRB3*02:02, report DRB1*11:01, DRB1*11:01 and DRB3*02:02, DRB3*02:02.
- If DRB1 does NOT support DRB3/4/5 homozygosity:
 - Enter the detected allele once.
 - Select “ABSENT” for the second field.
 - e.g. DRB1*01:01 with DRB1*11:01 is detected along with DRB3*02:02, for the DRB3/4/5 fields enter DRB3*02:02 and “ABSENT”.
- If tested but no DRB3/4/5 alleles detected:
 - Select “ABSENT” for both fields.
- If your lab does not test DRB3/4/5:
 - Leave fields blank.

G and P Group Reporting

- If a genotype cannot be resolved but alleles belong to the same G or P group, report the officially recognized WHO G or P group.
- Non-existent groups will be graded as **Discrepant**.
- Reference: <https://hla.alleles.org/>

Serologic Equivalents

After entering Class I and Class II results:

1. Click “Next” to go to the “Serologic Equivalents” tab.
2. If alleles appear with a drop-down list:
 - Select the appropriate serologic equivalent.
3. If no alleles appear:
 - No action is required as the serological equivalent is the same as the low-resolution molecular result.

Notes:

- Uncommon serologic equivalents (e.g., B*50:02 = B45) will not appear in the drop-down.
 - Add these in the Verification tab “Comments” field.
- For DRB3/4/5 null alleles:
 - Select “Not expressed.”
- For null alleles at other loci:
 - Indicate “not expressed” in the Verification tab “Comments” field.
- Serologic equivalents may be reported regardless of typing resolution selected.

Bw4/6 Entry

- If your laboratory does not report Bw4/Bw6 epitopes leave these fields blank.
- If reporting Bw4/Bw6:
- Enter results **based only on HLA-B antigens**
- Both fields must be completed or left blank

Examples:

- Both HLA-B antigens carry the Bw4 epitope → enter Bw4 in both fields.
- One HLA-B antigen carried Bw4 and the other Bw6 → enter Bw4 in the first field and Bw6 in the second field.
- One HLA-B antigen carries Bw6 and the other does not carry either Bw (e.g. B46, B73) → enter Bw6 and select ‘No Bw epitope’ for the second field.
- If both HLA-B antigens do not carry a Bw epitope (e.g. B46, B73) → select ‘No Bw epitope’ for both fields.

Final Step

- Once typing results entry and selection of associated Serologic equivalents (if present) and Bw4/Bw6 results in complete, click “Next.”
- Review entries in the “Verification” tab.
- Add any necessary comments.
- Click Save.

B27 RESULT ENTRY

For HT-B27 subscribers:

- Enter results in the separate “B27” section
- Confirm the test method for each sample ID
- Select if HLA-B27 was “positive” or “negative”
- Click “Save” at the bottom of the result entry screen

Please note: Laboratories may report only B27 results, if desired. To do so, remove all other HT analytes remove all other analytes (low resolution and high resolution HLA typing) by clicking the “X” next to the analyte name after clicking “Begin Survey”.

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Printing: Click the printer icon at the top of the page. Landscape orientation is recommended.

Laboratories are strongly encouraged to print results **IMMEDIATELY PRIOR TO SUBMISSION** and to review for accuracy and completeness of the entered and saved data.

Submitting Results: Results for ALL specimens must be entered and saved before survey submission.

To submit:

1. Click the orange “Submit Results” button (top and bottom right of the page)
2. Review results
3. Select ‘Yes’ to acknowledge data review
4. Click the orange “Submit”

Saved but unsubmitted entries will not automatically be submitted once the submission deadline has passed and will not be graded.

After successful submission:

- “SUBMISSION SUCCESSFUL” will be visible next to the survey name on the initial application screen and at the top and bottom of the Results Entry Screen.
- The orange “Submit data” button will no longer be visible.
- Access the submission date/time, cycle end date, and the “Submitted Results” PDF via the open book icon

Editing Submitted Results: Submitted results may be edited until the submission deadline.

To edit:

1. Click on the three dots [...] next to the specimen on the Data Results Entry Screen
2. Select “Edit” from the drop-down menu.
3. Edit results and re-review the serological equivalent assignment (if applicable)
4. Save results.
5. **Re-submit** results by clicking the orange “Submit Results” button (top and bottom right of the page)

If results are not re-submitted after the edit window has been opened ASHI will not be able to grade and score your results. Mistakes reported to the ASHI Proficiency Testing Program after the deadline has passed cannot be corrected.

Problems: If you experience problems entering or submitting results, please contact ASHI Headquarters.

Data Entry Mistakes: After submitting results, if you discover that you have made an error, you may re-enter the ASHI PT Data Center and edit your entries and re-submit your results up until the submission deadline. In general, mistakes reported to the ASHI Proficiency Testing Program after the deadline has passed cannot be corrected.

The ASHI PT Program cannot correct data entry errors that are detected by laboratories after a Discrepant result has been received.

Replacement Materials: Upon receipt of any sample, it is the responsibility of the receiving laboratory to inspect the shipment for damage. If a sample is received broken or in otherwise unsuitable condition, it is the responsibility of the receiving laboratory to notify the ASHI Central Office by phone or email within 24 hours of sample receipt. Documentation of the time of receipt of samples in the laboratory may be required.

For further details about replacement samples, please refer to the PT Operations Manual (link at top of document).

HT and B27 GRADING CRITERIA

Grading of HLA-B27 and HLA typing results requires a minimum of 10 participants, as well as 80% or higher consensus among participants. If fewer than 10 results are submitted for any challenge, those results will be ungraded. Performance grades are assigned separately for low resolution and high-resolution typing results. Each assigned antigen or allele is analyzed and graded separately but composite laboratory performance evaluation considers results for the complete HLA typing (combined results for HLA Class I and Class II) for each sample. For more detailed information regarding grading criteria, please refer to the ASHI PT Operations Manual under the section “Survey” and the appropriate survey name. A downloadable PDF version of the Manual is available by going to the PT resource page (<https://www.ashi-hla.org/page/PT>) or by using the link provided at the top of this document.

Contact Information for the ASHI Proficiency Testing Program:

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