



Recent Trends in the Evaluation of Analytical Biosimilarity

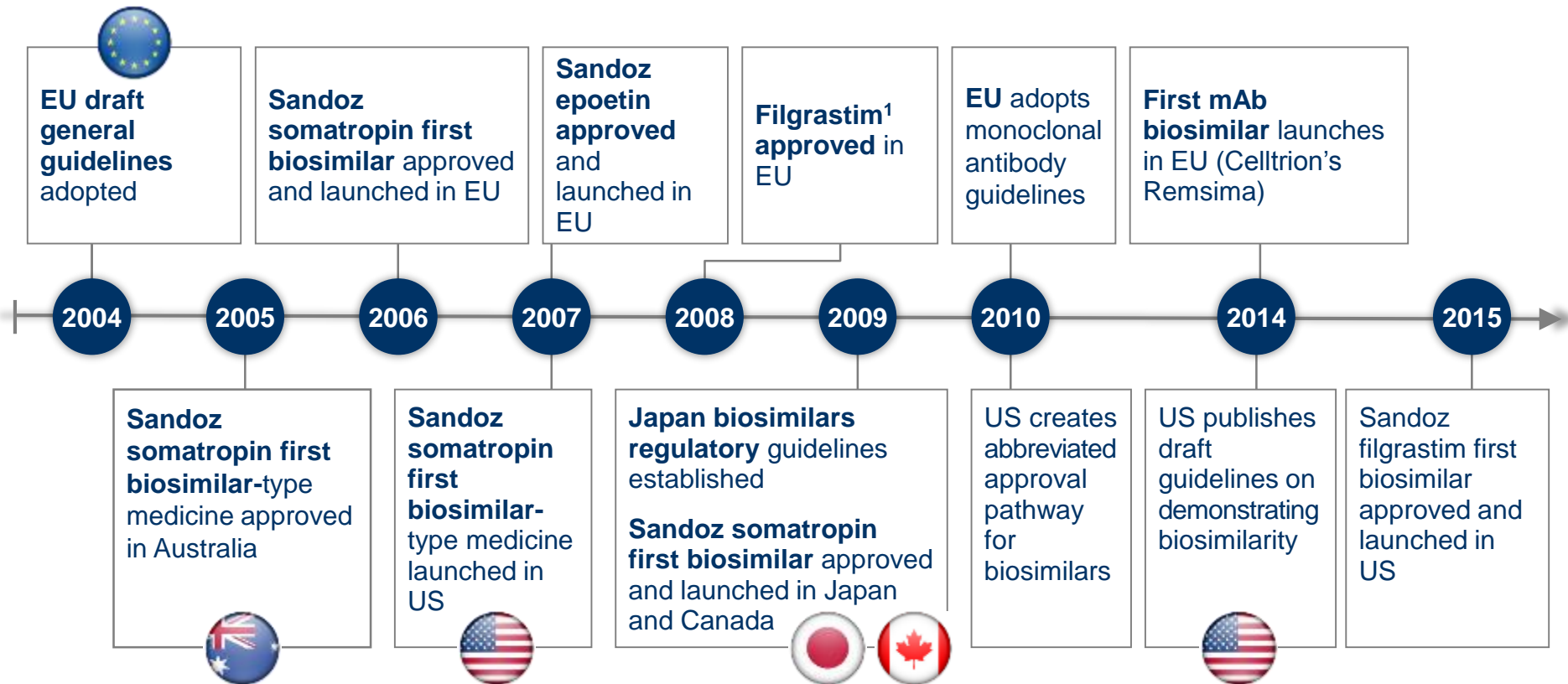
WCBP 2016, Washington D.C.

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Sandoz Biopharmaceuticals



Biosimilars are recognized around the world as safe and effective medicines



1 First competitor product (Sandoz product approved Feb 2009)

10 Years of Biosimilars...

... and evolving analytical technologies,
development concepts and regulatory sciences

Increases from 2005 to 2015:

Number of reference product batches*: x12

Number of analytical methods: x4

Number of quantitative readouts: x10

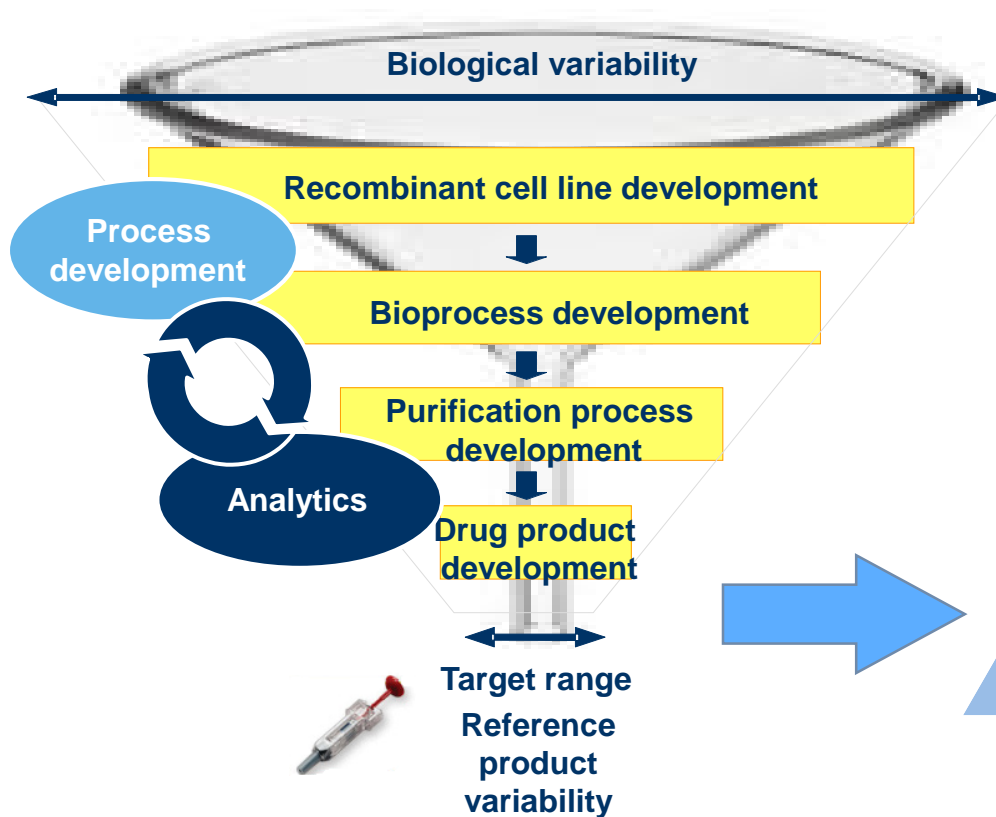
Pages of biosimilarity exercise: x7

Molecule size (mol. weight): x8

* as part of the comparability exercise

Biosimilars are systematically and iteratively developed to match the reference product

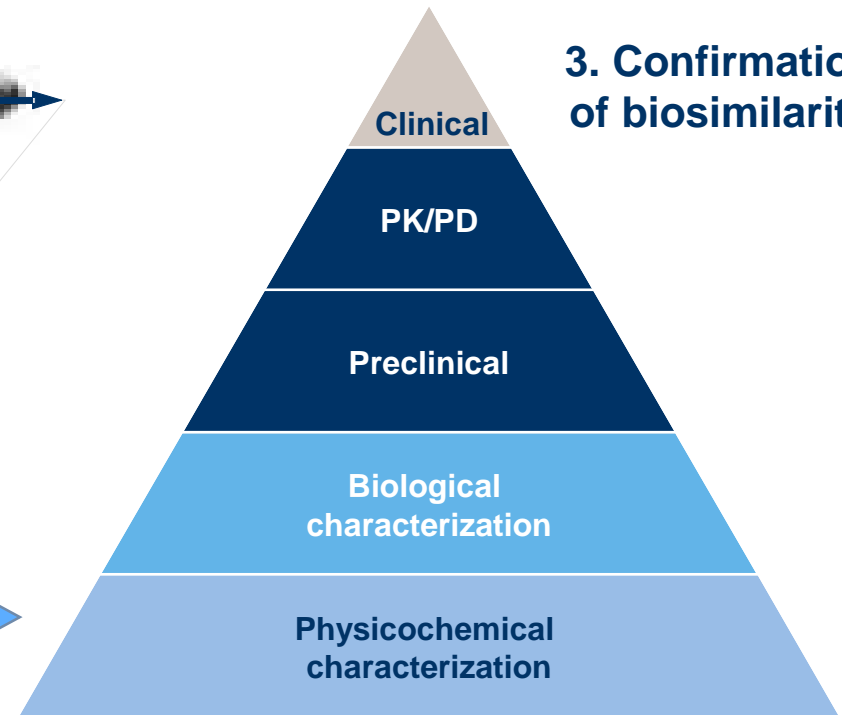
2. Target directed development



1. Target definition

No clinically relevant differences

3. Confirmation of biosimilarity

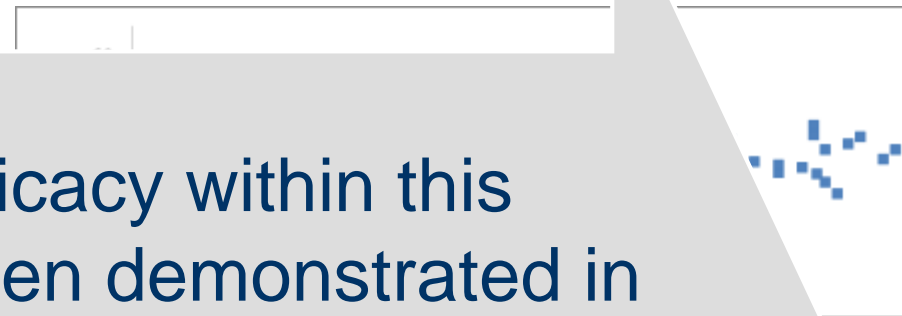


Variability is inherent in biologics

Batch-to-batch

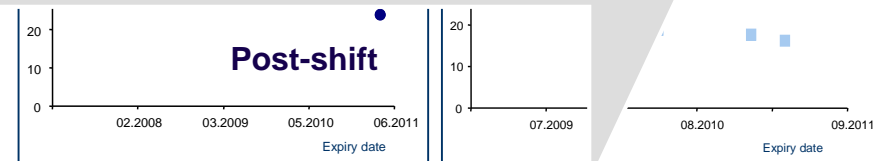
- Non-identity variability is a normal principle in biologics
- No batch-to-batch variability in the commercial mAB
- Variability in attributes sometimes significantly larger than batch-to-batch variability

Variability of major glycan variant in commercial mAB



Safety and efficacy within this variability have been demonstrated in clinical studies and by real-life experience with the reference product

- Manufacturing process improvements, scale up, etc
- Differences in attributes sometimes significantly larger than batch-to-batch variability



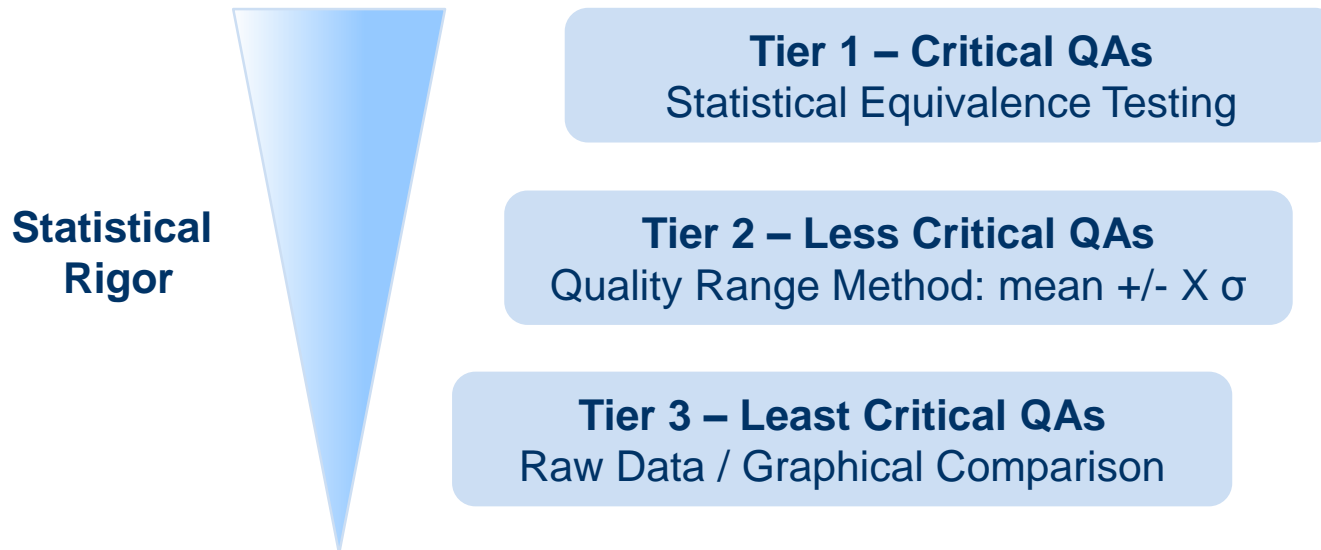
* M. Schiestl et al. Acceptable Changes in Quality Attributes of Glycosylated Biopharmaceutical; Nature Biotechnology (2011) 29:310

Considerations impacting biosimilarity evaluation

- The variability of the originator defines the goal posts for development
- Is there any difference between a
 - target range for development of a biosimilar
 - acceptance range for the biosimilarity exercise?
- Is every marketed batch from the originator defining acceptable quality with respect to its quality characteristics?
 - would a given quality characteristic of the originator lot be acceptable for a biosimilar lot?
- How to use the variability of the originator and the biosimilar to quantitatively assess for biosimilarity on a quality level
 - which statistical approach?
 - is statistics the deciding tool?

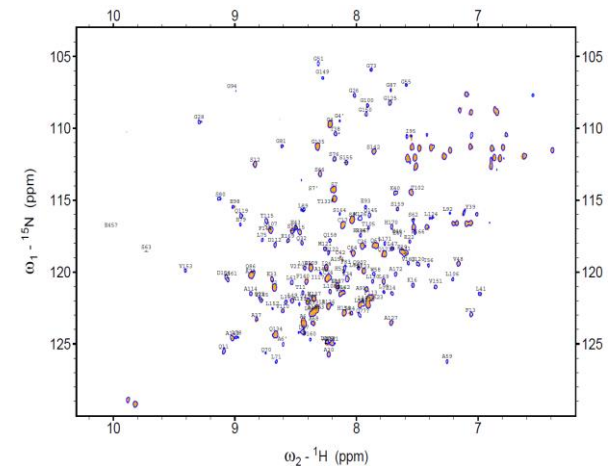
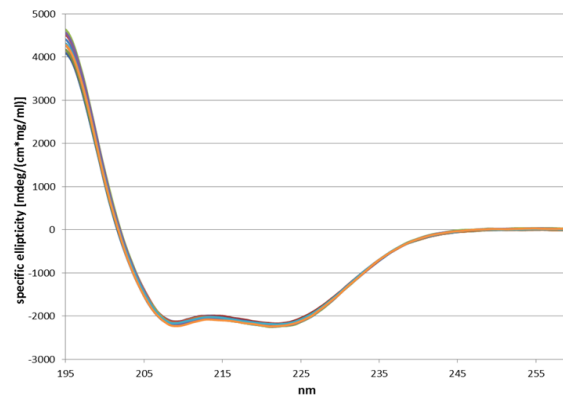
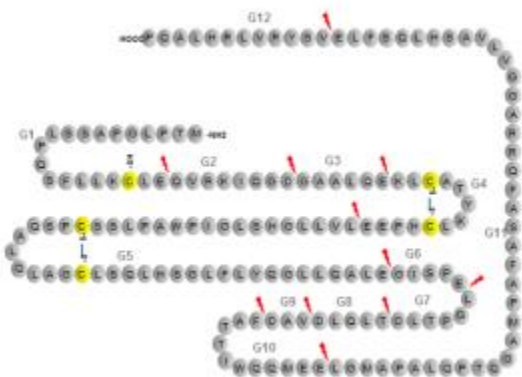
FDA's 3-Tier Approach

1. Evaluate the criticality of quality attributes
 - Impact on clinical performance
 - Degree of Uncertainty in Impact
2. Assign quality attributes to different tiers based on their criticality
3. Different statistical/quantitative approaches are applied to each tier



Not all quality attributes are evaluated best by statistical means

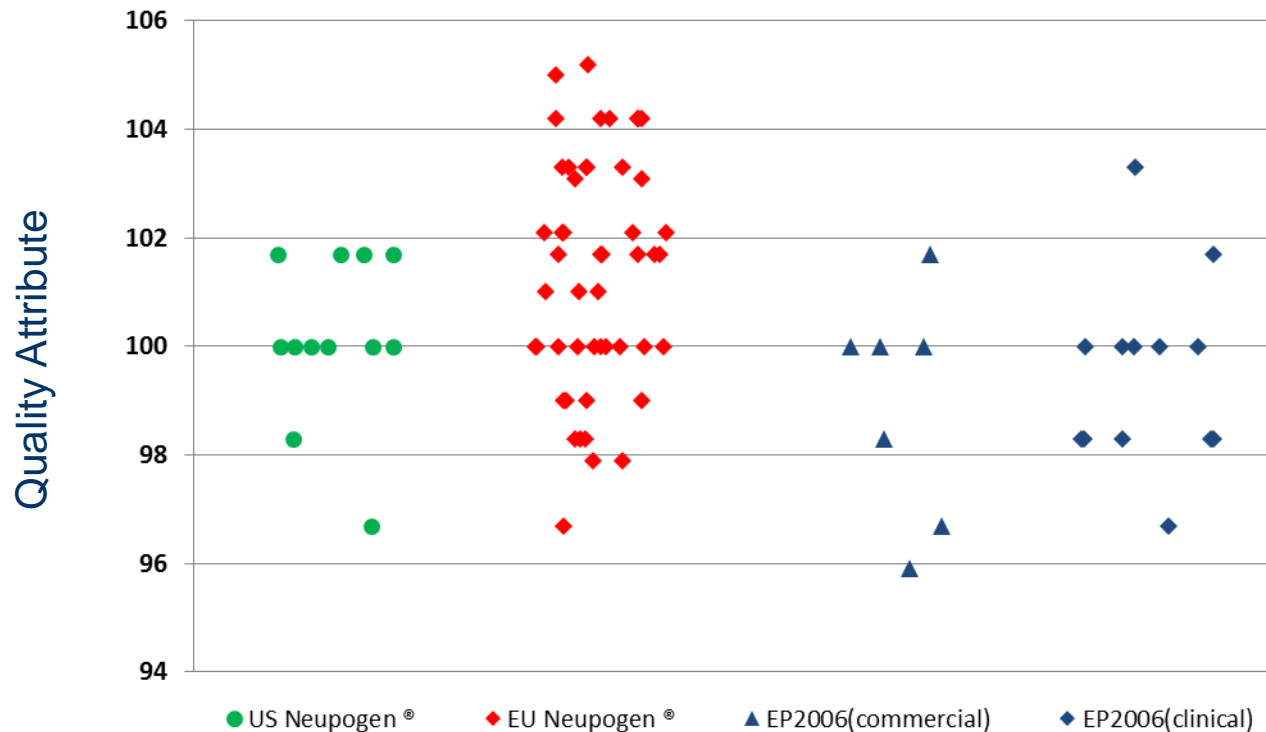
- Low criticality
- For some undesired quality attributes, „less than the maximum in reference product“ is better criterium than „equivalent“
 - Level of aggregates, deamidation, etc.
- For some quality attributes use of statistics less appropriate due to the nature of the data delivered by the particular analytical method



Source: Sandoz presentations for the January 7, 2015 Meeting of the Oncologic Advisory Committee

A graphical data comparison can already be very informative...

Comparing a biosimilar to its originator in different regions



Reference: Sandoz and FDA presentations for the January 7, 2015 Meeting of the Oncologic Advisory Committee

Tier 2 – Applying Quality Ranges

current practice based on reference product stdev

- Tier 2 testing is based on a quality range that depends on an estimate of the reference product standard deviation

Mean of reference product lots $\pm k \times \sigma_{RP}$

- A sufficient percentage of biosimilar batches (e.g. 90 %) required to fall into the quality range
- Multiplier k to be justified by sponsor (k may be 2,3,...)
 - difficult to find a scientific rationale for different multipliers for different quality attributes / analytical readouts other than criticality
 - k = 2 is too narrow to have a reasonable probability for two identical products (same μ and σ) to pass the criterion¹⁾
 - k = 3 is widely accepted as reasonable estimator of the realistic variability
 - common standard in statistical process control
 - „three-sigma rule of thumb“: „nearly all“ values within 3 sigma ²⁾

1) D. Weese & R. Burdick, IABS/FDA Statistical and Data Management Approaches for Biotechnology Drug Development, September 2015

2) Erik W. Grafarend, *Linear and Nonlinear Models: Fixed Effects, Random Effects, and Mixed Models*, Walter de Gruyter, 2006, p. 553

Statistical Equivalence Test for Protein Content

Protein content of the three product is statistically equivalent (mean values)

EP2006 vs US-Neupogen®
(-1.87, 0.15)



EP2006 vs EU-Neupogen®
(2.89, 0.85)



EU-Neupogen® vs US-Neupogen®
(0.27, 2.09)

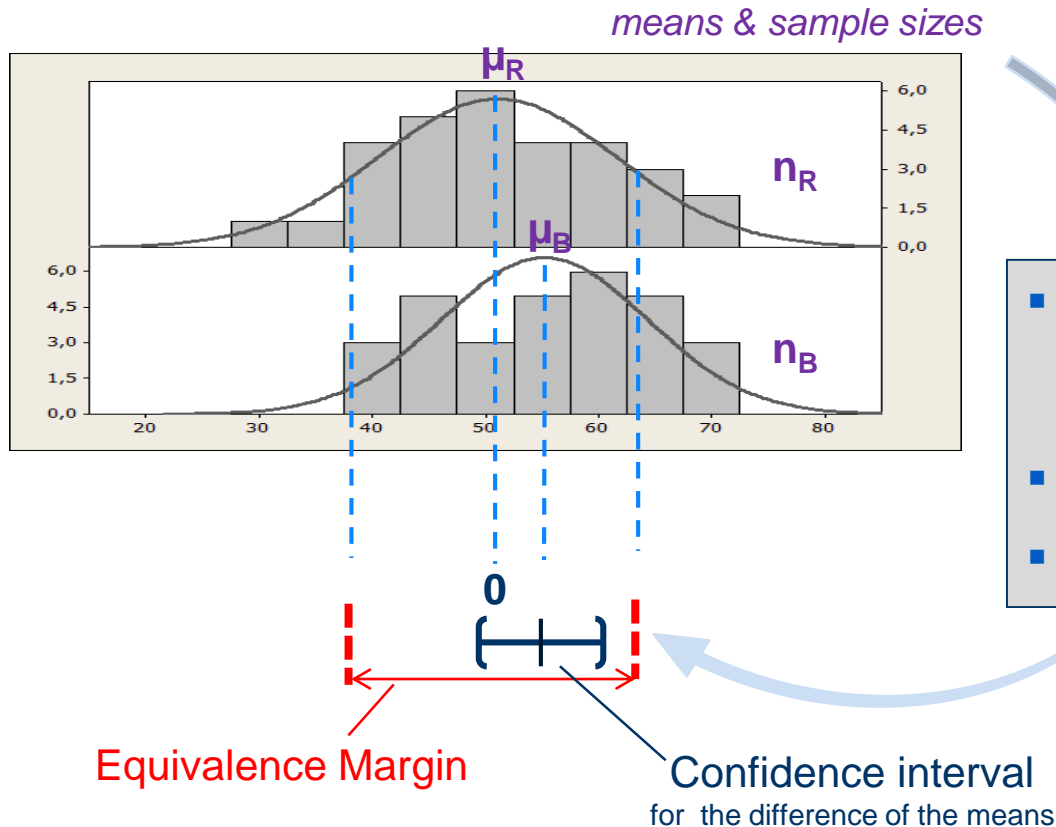


Results indicate that the products have the same strength and also support analytical similarity and analytical bridge

Source: FDA presentations for the January 7, 2015 Meeting of the Oncologic Advisory Committee

Equivalence testing for a practical difference in the means

Concluding equivalence by rejecting the null hypothesis $H: |\mu_R - \mu_B| \geq \delta$



The equivalence margin interrelates strongly to sample sizes, allowable difference, significance level and power

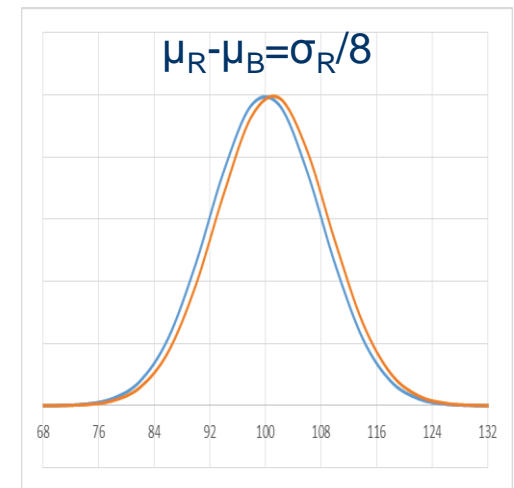
A reasonable choice of the equivalence margin is key for meaningful equivalence testing

- Determining the margin is challenging:
 - scientific justification usually not feasible
 - no standard statistical approach for determining the margin
- FDA's proposed equivalence margin is $1.5 \sigma_R$ independent of the sample size
 - easy to implement, no power calculation

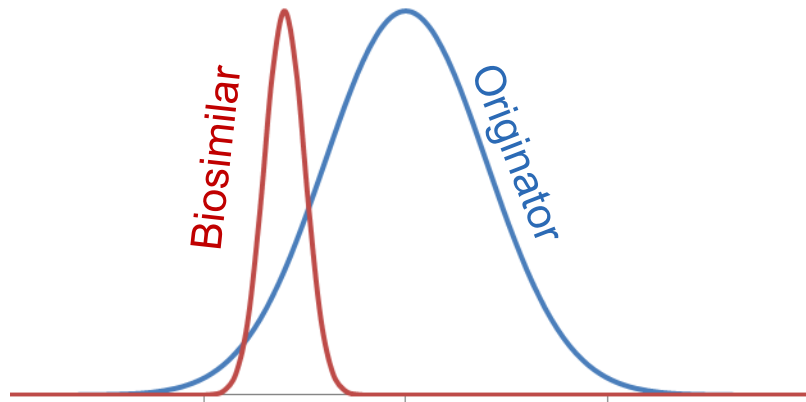
Background on the equiv. margin determination: 1)

- Sample sizes $n_R = n_B = 10$
- Difference of means between reference product and biosimilar $\mu_R - \mu_B = \sigma_R / 8$
- Probability to conclude „equivalent“ (power): 87 %

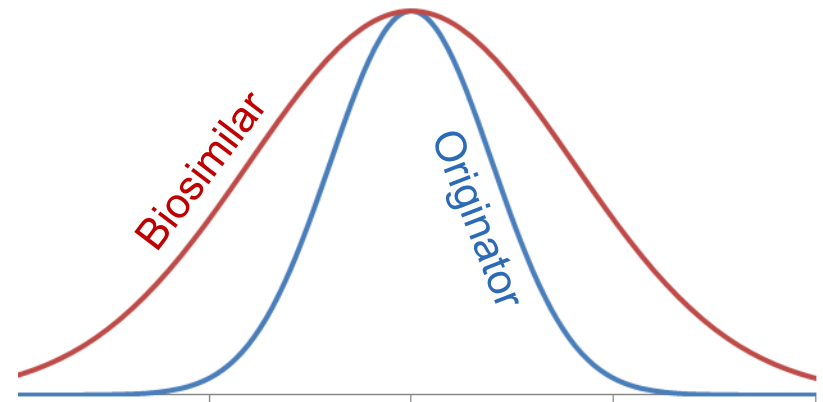
→ strong driver for large sample sizes for statistical (not necessarily scientific) reasons²⁾



The conceptual & theoretical implications of equivalence testing



All biosimilar batches are within variability of originator
means are different →
not equivalent

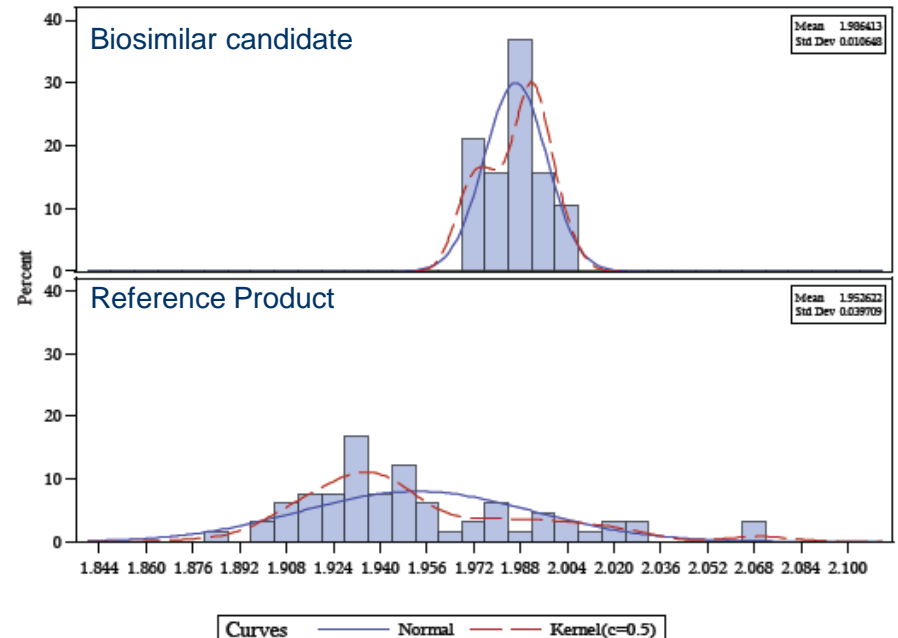
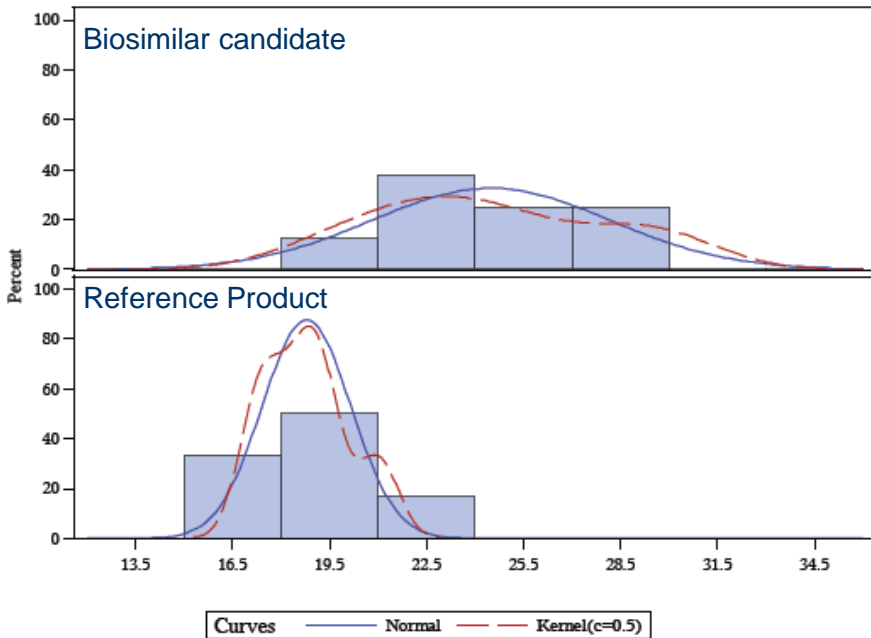


Some biosimilar batches are outside of the variability of originator
means are the same →
equivalent

The practical obstacles for statistics

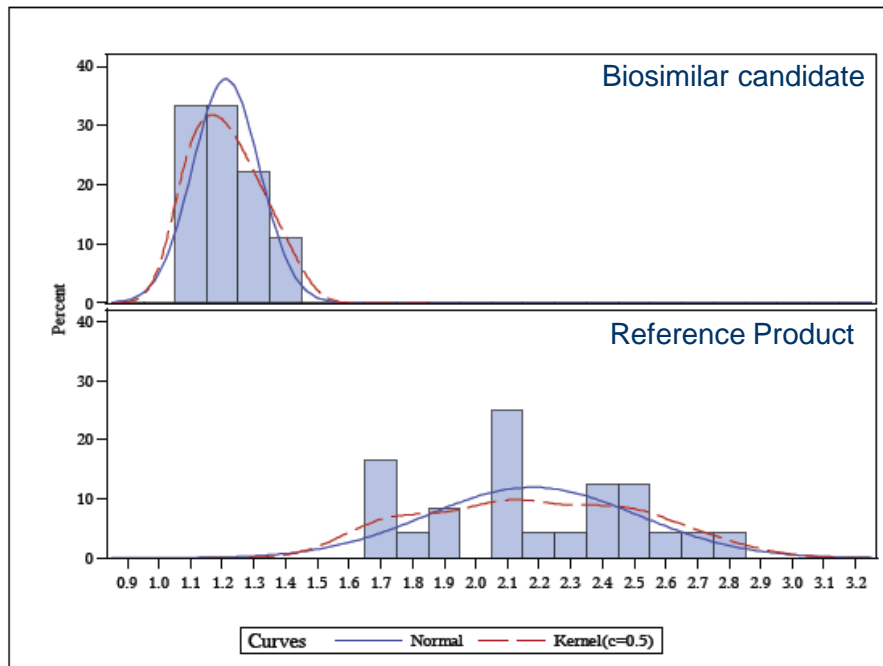
Very low sample sizes
& analytical variability

Non-normal distributions

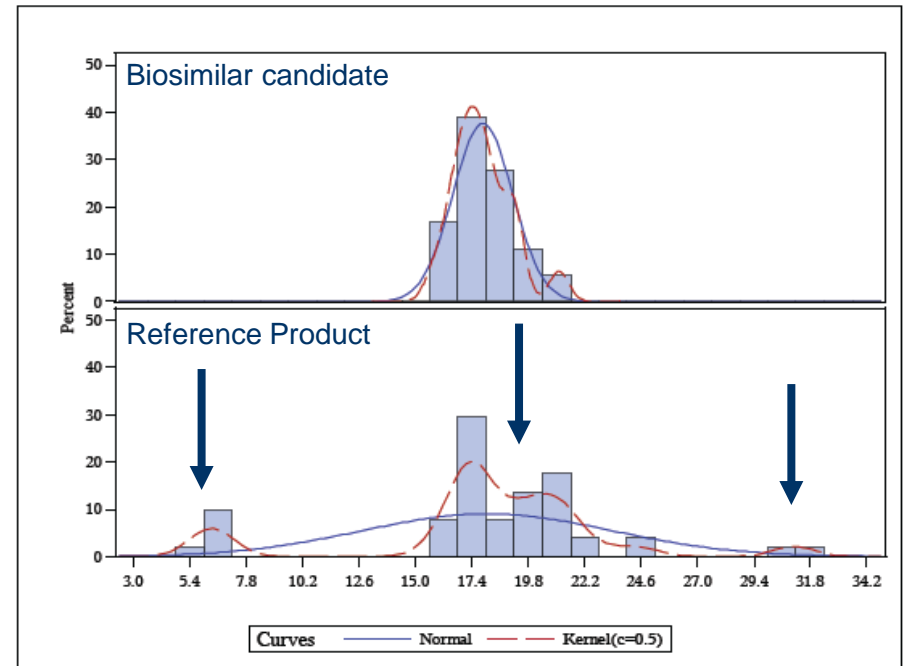


The practical obstacles for statistics

Undesirable quality attributes
(less is better)



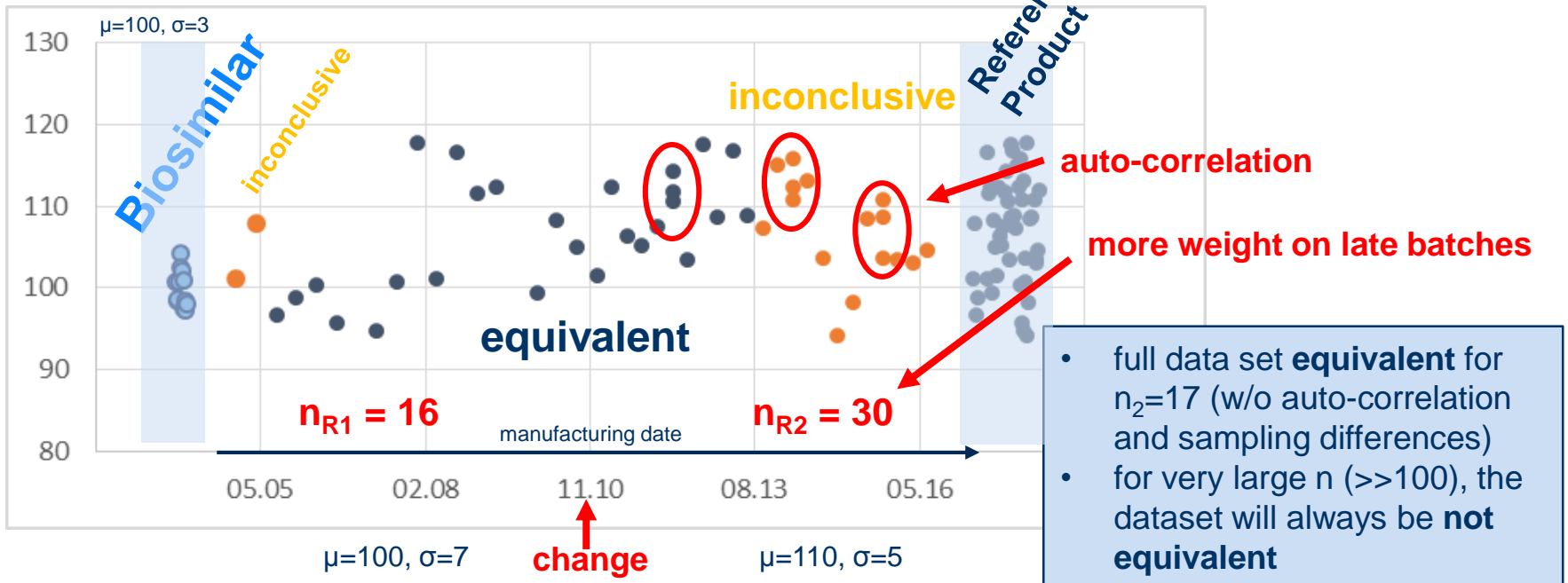
More than one reference product
population & „outliers“
due to manufacturing changes, long-term common
cause variability, or special cause variability



The pitfalls of diligently testing the reference product

Additional batches	Sampling differences	Auto-correlation
manufacturing change, long-term common cause variability	Different weight on different times	campaign production, 1 DS in several DP batches,...
→ shift in mean	→ shift in mean	→ decrease of stdev

simulated data



Implications of sampling reference product lots across many years

- Batch purchasing schedule (sampling) may impact the equivalence test
- If a biosimilar tests as equivalent – be cautious not to test it into inconclusive by additional reference product batches
- Equivalence testing does not allow for the definition of a useful development target

An inspirational comparison of the use of statistical tools for clinical studies vs. comparability

Clinical studies	CMC comparability
One primary endpoint	Multiple endpoints: quality attributes
Measure the physiological reaction after drug application	Measure the quality attributes of a given drug
Variability of the physiological processing Stratified random sampling	Variability of the manufacturing process Difficult to assure independent data
Acceptable margin for the primary endpoint based on clinical relevance	Acceptable margin based on scientific rationale -> Different for each quality attribute
<ul style="list-style-type: none"> ■ In the clinical evaluation, the predefinition of the endpoint and its related statistical evaluation is inevitable to mitigate the risk for bias ■ In a comparability exercise, the endpoints are already set by the CQA assessment -> no risk for bias in selecting the „wrong endpoint“ 	
Statistics required for final judgment	Statistics merely facilitator to describe the level of residual uncertainty and thus the level of justification needed in case of differences

Final thoughts...

- First be clear about your scientific question, then choose the statistical tool, **and be aware of the limitations**
- With carefully chosen statistical test parameters, all described tools are able to flag those quality attributes which need further evaluation
 - May speed up final evaluation if statistics is set up in the right way
 - However, the incremental knowledge gain is very little compared to a descriptive but critical raw data comparison
- Failure to pass a statistical test does not preclude similarity. It is a tool to decide which parameters have to be discussed/investigated in more detail.
- Statistics should not be a self-contained claim for biosimilarity on the quality level, it always should be just a contributor to the totality of evidence



Thanks for listening!

Thanks for contributing!

Colleagues at Mengeš, Schafteuau & Kundl, Oberhaching & Holzkirchen

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