



## CANINE DISTEMPER (CD)

ANIMAL GROUP AFFECTED	TRANSMISSION	CLINICAL SIGNS	FATAL DISEASE ?	TREATMENT	PREVENTION & CONTROL
Domestic and wild carnivore species, Javelinas, Tayassuidea	Aerosol or contact with oral, respiratory and ocular fluids, exudates and faeces	Depression, mucopurulent oculonasal exudates, CNS signs	Variable, between inapparent infections and high mortality	Modified live and dead virus vaccines, serum therapy (Stagloban)	<i>In houses</i>  <i>in zoos</i> Husbandry protocols, including adequate vaccination, quarantine and housing arrangements

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<b>Susceptible animal groups</b> Species in all families in the order Carnivora are susceptible to CDV (Canidae, Mustelidae, Procyonidae, Hyaenidae, Ursidae, Viverridae, and Felidae). Besides that even Javelinas and Tayassuidea are susceptible to CDV.	
<b>Causative organism</b> Canine distemper virus (CDV) is a negative-stranded RNA virus, size (100-250 nm) within the family <i>Paramyxoviridae</i> , subfamily <i>Paramyxovirinae</i> , and genus <i>Morbillivirus</i> . CDV is relatively fragile and quickly inactivated in the environment, by ultraviolet light, heat and drying and in common disinfectants.	
<b>Zoonotic potential</b> There has been concern that CDV could be associated with multiple sclerosis in humans, though this theory remains controversial.	
<b>Distribution</b> Animals: world-wide among wild and captive carnivores. Concern has been raised about the potential transmission of CDV from domestic dogs in or adjacent to wildlife reserves to native wildlife. This may have occurred in the case of free-ranging African wild dogs, lions and was thought possible in the case of giant pandas in China.	
<b>Transmission</b> Transmission of CDV is primarily by aerosol or contact with oral, respiratory, and ocular fluids and exudates containing the virus. It is shed from skin and in faeces and urine. Transplacental transmission has been documented in dogs. Close association between affected and susceptible animals is necessary due to the relative fragility of CDV in the environment. The virus may be shed for up to 90 days after infection. Suspected animals responsible for CDV transmission to zoo animals are mustelids, canids and felids. If dogs are allowed to enter zoos, they are able to transmit CD in cases where they did not obtain a proper immunoprophylaxis by vaccination.	
<b>Incubation period</b> Incubation period ranges from about 1 week to 1 month.	
<b>Clinical symptoms</b> Clinical presentation depends somewhat on the species; the classic signs are depression and mucopurulent oculonasal exudates. Fever, anorexia, vomiting, and diarrhoea frequently occur. Animals that recover from clinical CD or that have a prolonged illness may be in poor body condition. Central nervous system signs may	



be concurrent or follow systemic disease. Neurologic signs depend on the area of brain affected and include abnormal behaviour, convulsions, cerebellar and vestibular signs, paresis or paralysis and inco-ordination.
<b>Post mortem findings</b> Interstitial pneumonia to bronchopneumonia, catarrhal to haemorrhagic enteritis, and hyperkeratosis of the nose, lips, eyelids, ears, anus, and footpads may be present. Absolute lymphopenia is common. Less commonly observed lesions include damage of enamel, dentin, and tooth roots in animals that contact CDV prior to eruption of permanent teeth. An important diagnostic feature of CD is the presence of intracytoplasmic and intranuclear eosinophilic inclusion bodies in epithelia, neurons, and astroglia. Inclusion bodies are also often in gastric mucosa, enterocytes, pancreatic and biliary duct epithelium, and epithelium of the respiratory and urogenital tract.
<b>Diagnosis</b> Virus neutralisation test (NPLA) is the standard serologic test for antibodies against CDV. PCR also detects CDV in tissues. Immunohistochemistry is very useful for detection of CDV antigen in formalin-fixed, paraffin-embedded tissues.
<b>Material required for laboratory analysis</b> Sera, tissue (lung, spleen, brain); storage of tissue samples: -80°C; storage of serum samples: -20°C
<b>Relevant diagnostic laboratories</b> Dr. L. Haas, Institut für Virologie der Tierärztlichen Hochschule Hannover, Bünteweg 17, 30559 Hannover, Germany, Tel. +49-511 9538860, Fax +49-511 8568860 Dr. K. Frölich, Institut für Zoo- und Wildtierforschung, Alfred-Kowalke-Straße 17, 10315 Berlin, Germany Tel. +49-30 51 68-0, Fax +49-30 5126104
<b>Treatment</b> Wild animals that survive CD probably have lifelong immunity to subsequent infection by CDV. Modified live virus (MLV) vaccines, in species that can be safely vaccinated, induce long-life immunity. However, an inactivated vaccine is recommended in general. A serum therapy and the application of antibiotics is although recommended. To protect pantherides against CD a vaccination scheme involving both inactivated and attenuated live virus vaccine strains might be an option.
<b>Prevention and control in zoos</b> CD is one of the most important infectious diseases of carnivores in captivity and must be considered in design of husbandry protocols, including vaccination, quarantine, and housing arrangements. Because many threatened and endangered carnivores are susceptible to CDV, it must be considered in management plans and during restoration efforts. Cave: Common animal vaccines may be virulent for exotic carnivores.
<b>Suggested disinfectant for housing facilities</b> All virus disinfectants can be used.
<b>Notification</b>
<b>Guarantees required under EU Legislation</b>
<b>Guarantees required by EAZA Zoos</b>
<b>Measures required under the Animal Disease Surveillance Plan</b>
<b>Measures required for introducing animals from non-approved sources</b>
<b>Measures to be taken in case of disease outbreak or positive laboratory findings</b>
<b>Conditions for restoring disease-free status after an outbreak</b>
<b>Contacts for further information</b> <ul style="list-style-type: none"><li>• Dr. M. Appel, 13 Redwood Lane, Ithaca, NY 14850, USA, Email: mja13@cornell.edu</li><li>• Dr. L. Haas, Institut für Virologie der tierärztlichen Hochschule Hannover, Bünteweg 17, 30559 Hannover, Germany</li><li>• Prof. H. Lutz, Klinisches Labor, Tierspital Zürich, Winterthurerstraße 260, CH-8057 Zürich, Switzerland, Email: hanslutz@vetklinik.unizh.ch</li><li>• Dr. K. Frölich, Institute for Zoo and Wildlife Research, Alfred-Kowalke-Straße 17, 10315 Berlin, Germany</li></ul>

**References**

1. Alexander, K. A., and M. J. G. Appel. 1994. African wild dogs (*Lycaon pictus*) endangered by a canine distemper epizootic among domestic dogs near the Masai Mara National Reserve, Kenya. *J. Wildl. Dis.* 30: 481-485.
2. Appel, M. J. G., and D. S. Robson. 1973. A microneutralization test for canine distemper virus. *Am. J. Vet. Res.* 34: 1459-1463.
3. Appel, M. J. G. 1987. Canine distemper virus. *In: Appel, M. J. G. (ed.). Virus infections of carnivores.* Elsevier Science, Amsterdam. Pp. 133-159.
4. Budd, J. 1981. Distemper. *In: Davis, J. W., L. H. Karstad, and D. O. Trainer (eds.). Infectious diseases of wild mammals.* Iowa State University Press, Ames. Pp. 31-44.
5. Dubielzig, R. R. 1979. The effect of canine distemper virus on the ameloblastic layer of the developing tooth. *Vet. Pathol.* 16: 268-270.
6. Frölich, K., O. Czupalla, L. Haas, J. Hentschke, J. Dedek, and J. Fickel. 2000. Epizootiological investigations of canine distemper virus in free-ranging carnivores from Germany. *Vet. Microbiol.* 74: 283-292.
7. Greene, C. E., and M. J. Appel. 1998. Canine distemper. *In: Greene, C. E. (ed.). Infectious diseases of the dogs and cats.* W. B. Saunders, Philadelphia, Pennsylvania. Pp. 9-22.
8. Haas, L., H. Hofer, M. East, P. Wohlsein, B. Liess, and T. Barrett. 1996. Canine distemper virus infection in Serengeti spotted hyaenas. *Vet. Microbiol.* 49: 147-152.
9. Haas, L., W. Martens, I. Greiser-Wilke, L. Mamaev, T. Butina, D. Maack, and T. Barrett. 1997. Analysis of the haemagglutinin gene of current wild-type canine distemper virus isolates from Germany. *Virus Res.* 48: 165-171.
10. Krakowka, S., E. A. Hoover, A. Koestner, and K. Ketring. 1977. Experimental and natural occurring transplacental transmission of canine distemper virus. *Am. J. Vet. Res.* 38: 919-922.
11. Maack, D., M Böer, H.-P. Brandt, and B. Liess. 2000. Morbillivirus infections in German zoos: prevalence in carnivores and vaccination trials in pantherid cats. *EAZWV* 3: 47-53.
12. Maika, S. A. , X. Qui, T. He, and M. J. Appel. 1994. Serologic survey of giant pandas (*Ailuropoda melanoleuca*), and domestic dogs and cats in Wolong Reserve, China. *J. Wildl. Dis.* 30: 86-89.
13. Montali, R. J., C. R. Bartz, and M. Bush. 1987. Canine distemper virus. *In: Appel, M. J. G. (ed.). Virus infections of Carnivores.* Elsevier Science, Amsterdam. Pp. 437-443.
14. Williams, E. S., and E. T. Thorne. 1996. Infectious and parasitic diseases of captive carnivores, with special emphasis on the black-footed ferret (*Mustela nigripes*). *Rev. Sci. Tech. Off. Int. Epizoot.* 15: 91-114.
15. Williams, E. S. 2001. Canine Distemper. *In: Williams, E. S., and I. K. Barker (eds.). Infections and parasitic diseases of wild mammals.* Iowa State University Press, Ames. Pp. 50-59.