

# **In-Straw Dilution of Vitrified Bovine Embryos Produced In-Vitro**

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Commercial cryopreservation of bovine embryos has relied mainly on slow-cooling techniques. We have developed a simple, two step vitrification technique with in-straw dilution so that embryos can be transferred in the field. We studied factorial combinations of two equilibration concentrations of ethylene glycol (EG) (3.5 and 5 M), two thawing temperatures (20 and 37°C) and two post-thaw waiting times until culture (5 and 15 min). A total of 486 blastocysts sired by three bulls were obtained in vitro in six replicates. Briefly, oocytes were aspirated from 2-8 mm follicles from slaughterhouse ovaries, and matured, fertilized, and cultured in vitro with standard procedures. Vitrification was with 0.25-ml straws preloaded with a 1 cm column of D-HCDM (0.5 M galactose in Hepes-buffered holding medium (HCDM2)), then 0.5 cm air, and then 7 cm of D-HCDM. Embryos were transferred to 1 ml of V1-CDM (3.5 M or 5M ethylene glycol in HCDM2) for 3 min at 24°C. Next, embryos were moved in 1 µl into a 7 µl droplet of V2-CDM (7 M ethylene glycol, 0.5 M galactose, 18% w/v Ficoll 70 in HCDM2) at 24°C. In less than 1 min, the droplet containing the embryo was loaded, followed by 0.5 cm air and 1 cm of D-HCDM. Straws were heat-sealed and plunged vertically, sealed end first, into liquid nitrogen, covering the embryo; then the rest of the straw was slowly immersed. Straws were thawed in air (24°C) for 10 sec and then in water horizontally at 20 or 37°C until ice disappeared. Straws were gently shaken to mix the columns; then, after 5 or 15 min at 24 or 37°C, embryos were expelled and cultured in CDM2 + 5% FCS. Re-expansion and hatching rates were evaluated 48 h post thaw. Data (Table 1) were calculated as a percentage of non-vitrified controls for respective replicates (control means: re-expansion 86%; hatching 54%) and analyzed by ANOVA.

TABLE 1. Mean responses (LSM ± SE) of vitrified embryos (% of non-vitrified controls).

| Post-thawing<br>waiting time<br>(min) | Thawing<br>Temp. (°C) | <u>Re-expansion (%)</u>                      |      | <u>Hatching (%)</u> |       |
|---------------------------------------|-----------------------|----------------------------------------------|------|---------------------|-------|
|                                       |                       | <u>Equilibration concentration of EG (M)</u> |      |                     |       |
|                                       |                       | 5                                            | 3.5  | 5                   | 3.5   |
| 5                                     | 20                    | 84±10                                        | 86±6 | 49±12               | 79±16 |
|                                       | 37                    | 96±9                                         | 78±8 | 103±22              | 79±12 |
| 15                                    | 20                    | 71±10                                        | 64±4 | 48±17               | 42±6  |
|                                       | 37                    | 96±8                                         | 84±4 | 89±26               | 73±24 |

Main effect means for re-expansion were higher ( $P=0.08$ ) for 5 M (87 %) than 3.5 M (78%) EG for equilibration; also, re-expansion (88 and 76% of controls) and hatching (86 and 54 % of controls) were higher for 37 than 20°C ( $P<0.05$ ). There was no main effect for post-thaw waiting times for either response ( $P>0.1$ ), but there was an interaction between time and temperature for re-expansion ( $P<0.05$ ), indicating poor survival when embryos were thawed at 20°C and kept for 15 min. Equilibration with 5 M EG and thawing at 37 °C were optimal procedures, and embryos did not degrade when kept up to 15 min in straws thawed at 37 °C. Further studies including transfer of embryos to recipients are necessary to corroborate these results.

Key words: embryo, cryopreservation, vitrification, direct transfer.