

The diagnosis and treatment of endometritis in the mare: Yesterday and today

I.K.M. Liu ^{a,*}, M.H.T. Troedsson ^b

^a *Population Health and Reproduction, School of Veterinary Medicine, University of California, 1114 Tupper Hall, Davis, CA 95616, USA*

^b *College of Veterinary Medicine, University of Florida, Gainesville, FL, USA*

Abstract

Diagnosis and treatment of endometritis in the mare has been controversial and mostly empirical. The lack or inability of researchers to establish or develop a model that can serve as a standard or control makes this area of equine reproduction difficult to address scientifically. However, major advances have been made, particularly with the demonstration of the importance of uterine contractility in the elimination of bacteria, fluid, and inflammatory products from the uterus after breeding. This review provides a historical perspective of what has been done, and where we are now, in the approach to the diagnosis and therapy of endometritis in the mare.

Published by Elsevier Inc.

Keywords: Uterus; Uterine treatment; Endometritis; Equine; Mare

1. Introduction

Published reports in the literature on the diagnosis and treatment of endometritis in the mare are exhaustive and at times, controversial. Opinions vary considerably among individuals. Herein, the authors have formulated their own opinions based on the available literature, recognizing that these are not to be considered as generally accepted among all individuals.

Endometritis has been (over the past 40–50 y), and continues to be one of the major causes for infertility in the mare. Prior to the use of ultrasonography, methods for detecting endometritis in the mare ranged from seeing inflammation of the cervix with a speculum examination and observance of cervical and/or vaginal

discharge, to methods for culture [1], and cytological examinations [2]. The combined use of cytology and bacteriological culture as an aid to the diagnosis of endometritis in the mare was initially alluded to almost 20 y ago [3], and is currently the most commonly used procedure. Our methods for diagnosing endometritis in the mare have not changed (with the exception of instrumentation) since these early investigations, but with advancing knowledge of the pathophysiology of endometritis, treatments have changed. Therefore, it may be prudent to spend more of this discussion on the varying methods for treatment, rather than on the diagnosis of endometritis. Standard doses and protocols for the various treatments can be obtained from readily available texts and reviews. A summary of key events in the literature on the diagnosis and treatment of endometritis will be mentioned in this discussion, with an expectation that those not specifically mentioned should not be construed as unimportant.

* Corresponding author. Tel.: +1 530 752 1358.
E-mail address: ikliu@ucdavis.edu (I.K.M. Liu).

It was not until the late 1960s and early 1970s when it became clear that endometritis in the mare occurred in two forms: the acute phase and the chronic phase. In 1969, Hughes and Loy demonstrated that following the inoculation of *Streptococcus zooepidemicus* into the uterus of both young (maiden) and older (multiparous and barren) mares, a large inflammatory response was generated by the young mares, whereas little visual changes were noted in the older mares [4]. However, within a few days, the young mares no longer had evidence of inflammation or infection, whereas the older barren mares became chronically infected, with an obvious visual vaginal discharge. After spontaneously resolving the inflammation, the young mares were mated in the following cycle, and all became pregnant. From a practical viewpoint, this made a large difference in our diagnostic ability to determine which mares with endometritis required treatment and which do not. Hence, it is the persistence or the prolonged presence of endometritis that is responsible for subfertility/infertility in mares, and those that specifically require treatment.

2. Diagnosis and pathophysiology of endometritis

Prior to the use of ultrasonography, it was difficult to determine the presence, let alone the character and amount, of fluid present in the uterus of mares. Since its inception, the use of ultrasonography in reproduction has made a profound impact on our ability to detect the presence of intrauterine fluid in the mare, as well as its severity. This modality has increased our awareness and observations of endometritis, perhaps at earlier stages of uterine infection and compromised uterine clearance in mares. Retention of fluid in the uterus alone does not comprise a contribution to our diagnostic ability to detect endometritis. However, being able to assess the character of the fluid, and the volume of echodense fluid accumulation, gives us important information regarding the presence and severity of endometritis. In addition, we continue to use culture, as well as cytological and histological specimens, to assist us in the diagnosis of endometritis in the mare. These methods continue to be standard operating procedures in many broodmare practices, yesterday and today. An important complication observed over the years using standard practices (e.g. use of guarded swabs of the endometrium) of culture and cytology has been the presence of false-positive and false-negative results. For barren and subfertile mares, low-volume uterine flushes have been proposed as an alternative to the use of guarded swabs in

diagnosing endometritis in the mare. Although large-volume uterine flushes of the endometrium have been used in the identification of various proteins, bacteria and chemical components of the uterus [5–8], Ball et al. first reported the use of low-volume uterine flush and its potential inclusion as a more effective method for the identification of bacterial pathogens and inflammatory response, with an emphasis on the barren and subfertile mare [9]. Recent reports confirm the use of low-volume lavage in a large number of mares as a useful diagnostic method in detecting chronic endometritis [10]. Low-volume lavage, although more cumbersome than standard procedures used for obtaining cultures and cytology with guarded swabs, may prove valuable for conditions of subclinical endometritis in the chronically infected mare. Further improvements in obtaining more representative samples of endometrial contents have been suggested by John Steiner (personal communication, 2007) using a cervical brush adapted from instrumentation used for cervical cytology procedures in women. Clinicians having the opportunity to use this instrument (Minitube of America, Verona, WI, USA), indicate that this procedure has valuable potential as an alternative to currently used cotton-tipped swabs. Recently, Nielsen (from Denmark) compared and evaluated the endometrial swab and endometrial biopsy procedures for obtaining cytological and culture results for diagnosing endometritis in the mare [11]; in that study, endometrial biopsy was apparently a more sensitive and specific method in diagnosing endometritis in the mare than the use of endometrial guarded swabs. However, that study did not distinguish the differences between mares with acute or persistent endometritis. Since persistent endometritis (clinical and subclinical) is a more important issue to the practitioner, further investigations may be required to evaluate its effectiveness solely under these conditions.

It has been proposed that endometritis, both acute and chronic, is initiated by two distinctly separate origins, bacterial contaminants and sperm [12]. Only 10–15% of all brood mares develop a pathological persistent form of breeding-induced endometritis [13]. Attention should be given to identify and manage these mares appropriately in order to optimize reproductive efficiency. The inflammatory response to either antigen is similar, in that an acute or persistent inflammatory response may ensue. Hence, treatment strategies may be somewhat different, based on the origin of the disease. Although other currently unknown factors may also be involved in the development of persistent endometritis, it is clear that the major factor influencing the retention of endometrial inflammatory responses, whether they

are bacterial or sperm-induced, is based on delayed uterine clearance as a result of compromised myometrial function [14–16]

For many years, investigators hypothesized the importance and studied the role of immunoglobulins and polymorphonuclear leukocytes as they relate to persistent and recurring endometritis. Although evidence was provided that immunoglobulins and PMNs played key roles in persistent uterine infections, a few notable studies demonstrated that compromised uterine clearance and myometrial contractility played a major, and perhaps a more important, role in the pathophysiology of mares affected with chronic uterine infections. Using chromium⁵¹ labeled microspheres, it was shown that mares potentially susceptible to chronic uterine infections failed to clear the microspheres adequately when compared with mares with normal uteri [14]. Using electromyographic recordings, these investigators further provided direct evidence that the cause for poor uterine clearance was due to compromised uterine contractility [15]. Similarly, LeBlanc et al., in their elegant study using radioactive colloids and scintigraphy, also demonstrated that mares with histories of chronic uterine infection have compromised uterine clearance [16]. These studies made a strong impact on our current treatment strategies of persistent endometritis in the mare. It is generally agreed among scientists and practitioners that if a mare is unable to spontaneously eliminate an infection in 4 d, she should be considered as being affected with persistent endometritis. It is our impression that if a mare is unable to spontaneously eliminate an infection after 2 d, she is either persistently infected, or is on her way of becoming persistently infected. This impression is supported by observations on uterine clearance of bacteria in normal mares [17].

3. Treatment strategies for endometritis

That previously acceptable pregnancy rates in most broodmare operations were within the 50–60% range is difficult to perceive in the context of current expectations. Today, acceptable ranges are 75–85%, with 90–100% ranges not being unusual. This increase in rates can be attributable to increased knowledge of the pathophysiology of endometritis and the infertile mare, better instrumentation, and more efficient and effective treatment strategies. A common goal for any broodmare practitioner is to get the mare pregnant. The single-most factor that has contributed to our increase in pregnancy rates has been the Caslick procedure. Historically, this was a fundamental requirement for most barren mares,

and it continues to be a major contributing factor to successfully achieving pregnancy.

When isolation of microbes from the uterus and sensitivity tests for antimicrobials were in vogue, extensive and repetitive use of antimicrobials was infused into the uterus of mares with endometritis. The use of antimicrobial intrauterine infusions is beneficial in cases of known bacterial growth causing endometritis and is currently used by many practitioners, including its use empirically as a single post-ovulation treatment in mares. Although the mechanism is not known, clinical impressions suggest that repetitive, prolonged and excessive use of antimicrobials appear to be associated with an increase in the incidence of fungal endometritis in mares, and that an increase in this condition was apparent in the 1980s and early 1990s. It is one of the authors' (IKL) belief that prolonged antimicrobial infusion of the uterus has not been used as extensively as it has been over the past 10 y, and concurrently, there has been an apparent decrease in fungal endometritis. Several anti-fungal agents including polyene antibiotics and azole derivatives are available and can be used as treatment strategies. An excellent review of equine fungal endometritis was reported by Dascanio et al., including treatment strategies and recommended doses [18]. Lufenuron[®], a benzoylphenyl urea derivative, has also been tested on a limited number of mares with fungal endometritis. Lufenuron[®] is believed to have an inhibitor effect on chitin production of fungal elements. Although the results appear encouraging, further investigations are required to evaluate its long-term effectiveness in the treatment of fungal endometritis in the mare [19].

Commonly used antimicrobial and anti-fungal drugs and dosages for treatment of endometritis in the mare are provided in the Manual of Equine Reproduction (edited by T. Blanchard, D. Varner, J. Schumacher, C. Love, S. Brinsco and S. Rigby) [20]. Included is a list of commonly used disinfectants that have been diluted and used as an alternative for antimicrobial treatments. Although commonly used among practitioners, very little is known about the efficacy of using diluted disinfectants when compared with antimicrobials.

Numerous agents have been used as chemical curettage for treatment of endometritis in the mare. Included are dimethyl sulfoxide (DMSO), hydrogen peroxide (H₂O₂), magnesium sulfate (MgSO₄), *Streptococcus* filtrate, kerosene, and dilute disinfectants (0.2% povidine-iodine solution). Available reports, anecdotal or in the literature, suggest that there is a beneficial effect in the treatment of chronic endometritis following its use. Although these agents represent a

variety of chemical components, they have a common denominator of inducing a strong inflammatory response when exposed to the endometrium. Hence, it is the authors' opinion (not based on scientifically meritorious investigations) that the limited success of these agents, when used as treatments for persistent endometritis, can be primarily attributed to the induction of a strong inflammatory response and inciting increased myometrial contractility and uterine clearance. Clearly, many of these agents may also have bactericidal effects. Furthermore, their use is generally reserved for barren mares in which no other treatments have been successful. Regardless of how effective the bactericidal effect, uterine contractility must be optimal for successful treatment to occur. Mechanical curettage is seldom used as a treatment for chronic endometritis in the mare in today's practices. Its use is perceived to be rather invasive, the effectiveness of the procedure has been questioned, and it is more than likely that chemical curettage has replaced its use.

Autologous/heterologous plasma and colostrum have also been used to treat endometritis [21,22]. Although colostrum is rarely used as a treatment regime today, its intended use as treatment for endometritis was to enhance the immunoglobulin population in the affected uterus, thereby assisting the immune response in its antimicrobial action. With increased knowledge regarding the pathophysiology of endometritis in the mare, although immunoglobulins do play a role in uterine defense, their role is presumed to be minor when compared with uterine clearance.

The infusion of autologous or heterologous plasma as a treatment strategy for endometritis is still currently being used in some practices. Its intended use and proposed success as treatment for endometritis was to enhance phagocytosis of bacteria by supplementation of opsonins (complement) present in the exogenous serum. Regardless of how effective the antimicrobial effect, uterine clearance and optimal uterine contractility are necessary ingredients for a successful outcome. Therefore, it may be presumptive to suggest that the successful treatments attributable to the infusion of colostrum and/or plasma may be related to a concomitant increase in uterine contractility and subsequent uterine clearance.

Supplementation of separated PMNs, fresh and cryopreserved, have also been reported in the literature and proposed as a successful treatment for persistent endometritis in the mare [23–25]. We are not aware of anyone using these treatment regimes in the field at this writing and presumably, these treatments have not gained popularity as yet in North America.

Treatment strategies have varied considerably among practitioners over the years. Increasing knowledge of the pathophysiology of persistent uterine infections and in particular, uterine lavage with isotonic saline and the accompanying use of ecbolics, has made the strongest contribution to the reduction of persistent endometritis and increased pregnancy rates in mares more than any other current treatment strategy in recent years. Uterine lavage was first used and described by Russian veterinarians as early as 1938 [26]. It was reported as an excellent means of eliminating *Escherichia coli* and *S. zooepidemicus* bacterial infections. Confirmation of uterine lavage as an effective treatment strategy for chronic endometritis was followed-up by Dr. Varadin from Yugoslavia, who reported in English, its successful use in mares with endometritis at the first International Symposium on Equine Reproduction in 1975 [27]. Subsequently, its use in North America has increased dramatically, based on procedural modifications, experiences and the promising results obtained from this procedure. An important contribution to the use of uterine lavage is the study by Brinsko et al., who reported that lavaging a mare 4 h after mating would not have any adverse effect on pregnancy in the mare [28]. This study gives the practitioner substantial flexibility in the timing of treatment of mares with bacterial- and/or sperm-induced recurrent endometritis and delayed uterine clearance. The use of ecbolics (oxytocin and prostaglandin), in conjunction with uterine lavage, was effective in enhancing uterine contraction and uterine clearance [16,29]. However, failure of pregnancy may result from the use of PGF_{2α} if administered during the periovulatory period (2–4 d) after ovulation [30,31]. Although other similar studies do not share this viewpoint [32], it is generally agreed that PGF_{2α} treatment 1–2 d after ovulation, causes a transient decrease in peripheral progesterone concentrations, followed by a rebound effect. Although controversy continues regarding the effects of prostaglandin on pregnancy when used during the periovulatory period, further investigations will be required to resolve this issue. Currently, oxytocin (an alternative ecboic) is suggested for post-ovulation treatments in mares with fluid retention. Regardless, it is clear that fluid remaining in the uterus after ovulation does not provide an optimal uterine environment, and its removal is mandatory to optimize embryo survival.

Immune stimulants have been promoted in recent years as a treatment for persistent endometritis in the mare [33]. There are at least two that are commercially available to the practitioner. One is a cell-mediated immunity stimulator, derived from inactivated *Propio-*

nibacterium acnes. The other, a *Mycobacterium* cell-wall extract, proclaims to normalize uterine inflammatory response, uterine involution and bacterial reduction in the uterus by a non-specific immunological response during endometritis in the mare. Despite preliminary evidence that the immune enhancers may be a useful treatment strategy for persistent endometritis in the mare, their efficacy requires further documentation in the field, as well as in established controlled studies.

Other less commonly used treatment strategies have recently been proposed. The use of corticosteroids in mares with excessive inflammation in response to breeding has been suggested [34]. The authors administered acetate 9- α -prednisolone (0.1 mg/kg) twice daily during estrus, starting when a follicle >35 mm was detected and ending when ovulation was confirmed. Preliminary results were encouraging, and further research is needed to clarify the mechanism of action for this treatment alternative.

Electro-acupuncture has also been used clinically to increase uterine contractility in mares with delayed uterine clearance. Anecdotal reports are encouraging, and research is needed to confirm the efficacy of this treatment alternative.

Finally, it is important for the clinician to remember that a transient inflammatory response to semen is normal, and required for normal fertility. Post-breeding treatments of these mares will most likely not improve fertility, but may even cause further contamination and interfere with pregnancy.

References

- [1] Dimock WW, Edwards PR. The pathology and bacteriology of the reproductive organs of mares in relation to sterility. *Bull Ky Agric Exp Stn* 1928;286.
- [2] Knudsen O. Endometrial cytology as a diagnostic aid in mares. *Cornell Vet* 1964;54:415–22.
- [3] Shin SJ, Lein D, Aronson AL, Nusbaum SR. The bacteriological culture of equine uterine contents, in-vitro sensitivity of organisms isolated and interpretation. *J Reprod Fertil Suppl* 1979;29:307–15.
- [4] Hughes JP, Loy RG. Investigations on the effect on intrauterine inoculation of *Streptococcus zooepidemicus* in the mare. *Proc Am Assoc Equine Pract* 1969;15:289–92.
- [5] Williamson P, Dunning A, O'Connor J, Penhale WJ. Immunoglobulin levels, protein concentrations and alkaline phosphatase activity in uterine flushings from mares with endometritis. *Theriogenology* 1983;19:441–8.
- [6] Stremienski PJ, Kenney RM. Effect of stage of cycle, sampling frequency and recovery of micro-organisms on total protein content of mare uterine flushings. *J Reprod Fertil* 1984;70:327–32.
- [7] Slusher SH, Freeman KP, Roszel JF. Eosinophils in equine uterine cytology and histology specimens. *J Am Vet Med Assoc* 1984;184:665–9.
- [8] El Sanousi SM, El Tayeb, Amna B. Bacteria isolated from uterine washings from mares in the Sudan. *Equine Vet J* 1979;11:219–22.
- [9] Ball BA, Shin SJ, Patten VH, Lein DH, Woods GL. Use of a low-volume uterine flush for microbiologic and cytological examination of the mare's endometrium. *Theriogenology* 1988;29:1269–83.
- [10] LeBlanc MM, Magsig J, Stromberg AJ. Use of a low-volume uterine flush for diagnosing endometritis in chronically infertile mares. *Theriogenology* 2007;68:403–12.
- [11] Nielsen JM. Endometritis in the mare: a diagnostic study comparing cultures from swab and biopsy. *Theriogenology* 2005;64:510–8.
- [12] Troedsson MHT. Breeding-induced endometritis in the mare. In Carnevale EM, editor. *Vet Clin North Am: Equine Pract* 2006;22:705–12.
- [13] Zent WW, Troedsson MHT. Postbreeding uterine fluid accumulation in a normal population of thoroughbred mares: a field study. In: *Proc 44th Ann Conv Am Assoc Equine Pract*. 1998. p. 64–5.
- [14] Troedsson MHT, Liu IKM. Uterine clearance of non-antigenic markers (^{51}Cr) in response to a bacterial challenge in mares potentially susceptible to chronic uterine infection. *J Reprod Fertil Suppl* 1991;44:283–8.
- [15] Troedsson MHT, Liu IKM, Ing M, Pascoe J, Thurmond M. Multiple site electromyography recordings of uterine activity following an intrauterine bacterial challenge in mares susceptible and resistant to chronic uterine infection. *J Reprod Fertil* 1993;99:307–13.
- [16] LeBlanc MM, Neuwirth L, Asbury AC, Mauragis D, Tran T, Klapstein E. Scintigraphic measurement of uterine clearance in normal mares and mares with recurrent endometritis. *Equine Vet J* 1994;26:279–84.
- [17] Katila T. Onset and duration of uterine inflammatory response of mares after insemination with fresh semen. *Biol Reprod Monogr* 1: *Equine Reprod VI*; 1995: 515–7.
- [18] Dascanio JJ, Schweizer C, Ley WB. Equine fungal endometritis. *Equine Vet Ed* 2001;13:324–9.
- [19] Hess MB, Parker NA, Purswell BJ, Dascanio JJ. *J Am Vet Med Assoc* 2002;221:266–7.
- [20] Blanchard TL, Varner DD, Schumacher J, Love CC, Brinsko SP, Rigby SL, editors. *Manual of equine reproduction*. 2nd ed., Mosby Inc.; 2003.
- [21] Asbury AC. Uterine defense mechanisms in the mare: the use of intrauterine plasma in the management of endometritis. *Theriogenology* 1984;21:387–93.
- [22] Dewes DF. Preliminary observations on the use of colostrum as an uterine infusion in thoroughbred mares. *N Z Vet J* 1980;28:7–8.
- [23] Zerbe H, Castilho LFF, Engelke F, Mattos RC, Schuberth HJ, Klug E, et al. Isolation and cryopreservation of functionally competent equine leukocytes. *J Vet Med* 2003;50:179–84.
- [24] Mattos RC, Castilho LFF, Malschitzky E, Mattos R, Gregory RM. Treatment of endometritis of experimentally infected mares. I. Effect of lavage, plasma and leukocytes on uterine bacterial clearance. *Arq Fac Vet – UFRGS* 1999;27:49–60.
- [25] Mattos RC, Neves AP, Gregory RM, Gomes MJS, Mattos R, Meirelles LS, et al. Treatment of endometritis of experimentally infected mares. II. Effect for ecobolic drugs and plasma enriched with leukocytes on uterine bacterial clearance. *Arq Fac Vet – UFRGS* 1999;27:61–72.
- [26] Zivotkov HI, Racializacija. I perestrojka slucki iiskustvenogo osemenenija kobil na novih proizvodstvenik inaucnih obosnovanijah. *Seljhovgiz, Moscow*; 1938.

- [27] Varadin M, Endometritis. A common cause of infection in mares. J Reprod Fertil Suppl 1975;23:353–6.
- [28] Brinsko SP, Varner DD, Blanchard TL. The effect of uterine lavage performed four hours post-insemination on pregnancy rates in mares. Theriogenology 1991;35:1111–9.
- [29] Troedsson MHT, Liu IKM, Ing M, Pascoe J. Smooth muscle electrical activity in the oviduct and the effect of oxytocin PGF₂a and PGE₂ on the myometrium and oviduct of the cycling mare. Biol Reprod Monogr 1: Equine Reprod 1995; 439–52.
- [30] Troedsson MHT, Ababneh MM, Ohlgren AF, Madill S, Vetscher N, Gregas M. Effect of periovulatory prostaglandin F₂ α on pregnancy rates and luteal function in the mare. Theriogenology 2001;55:1891–9.
- [31] Brendemuehl JP. Effect of oxytocin and cloprostenol on luteal formation, function and pregnancy rates in mares. Theriogenology 2002;58:623–6.
- [32] Nie GJ, Johnson KE, Wenzel JGW, Braden TD. Effect of periovulatory ecbolics on luteal function and fertility. Theriogenology 2002;58:461–3.
- [33] Rohrbach B, Sheerin P, Steiner J, Matthews P, Cantrell C, Dodds L. Use of *Propionibacterium acnes* as adjunct therapy in treatment of persistent endometritis in the broodmare. Anim Reprod Sci 2006;94:259–60.
- [34] Dell'Aqua Jr JA, Papa FO, Lopes MD, Alvarenga MA, Macedo LP, Neves neto JR, et al. Fertility rates with equine frozen semen after modulation of inflammatory uterine response. In: Proc 15th Int Cong Anim Reprod. 2004. p. 391 (abstract).